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| | | | 1634 | |

DATE MAILED: 07/07/2006

Please find below and/or attached an Office communication concerning this application or proceeding.

Office Action Summary

Application No.

10/738,546

Applicant(s)

WHEELER ET AL.

Examiner

BJ Forman

Art Unit

1634

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --

Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☐ Responsive to communication(s) filed on ____.
- 2a) ☐ This action is **FINAL**. 2b) ☒ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 1-26 is/are pending in the application.
- 4a) Of the above claim(s) ____ is/are withdrawn from consideration.
- 5) ☐ Claim(s) ____ is/are allowed.
- 6) ☒ Claim(s) 1-26 is/are rejected.
- 7) ☐ Claim(s) ____ is/are objected to.
- 8) ☐ Claim(s) ____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☒ The specification is objected to by the Examiner.
- 10) ☒ The drawing(s) filed on 16 December 2003 is/are: a) ☒ accepted or b) ☐ objected to by the Examiner.
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All b) ☐ Some * c) ☐ None of:
- ☐ Certified copies of the priority documents have been received.
 - ☐ Certified copies of the priority documents have been received in Application No. ____.
 - ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

* See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

- ☐ Notice of References Cited (PTO-892)
- ☐ Notice of Draftsperson's Patent Drawing Review (PTO-948)
- ☒ Information Disclosure Statement(s) (PTO-1449 or PTO/SB/08)
Paper No(s)/Mail Date ____.
- ☐ Interview Summary (PTO-413)
Paper No(s)/Mail Date. ____.
- ☐ Notice of Informal Patent Application (PTO-152)
- ☐ Other: ____.

DETAILED ACTION

Specification

1. The title of the invention is not descriptive. A new title is required that is clearly indicative of the invention to which the claims are directed.

The following title is suggested: METHOD FOR DESIGNING PROBE ARRAYS.

Priority

2. The first paragraph of the specification identifies numerous applications as "related". Applicant is reminded that for benefit claims under 35 U.S.C. 120, 121, or 365(c), the reference must include the relationship (i.e., continuation, divisional, or continuation-in-part) of all nonprovisional applications. If the application is a utility or plant application filed under 35 U.S.C. 111(a) on or after November 29, 2000, the specific reference to the prior application must be submitted during the pendency of the application and within the later of four months from the actual filing date of the application or sixteen months from the filing date of the prior application (See 37 CFR 1.78(a)(2)(ii) and (a)(5)(ii)).

Claim Rejections - 35 USC § 101

3. 35 U.S.C. 101 reads as follows:

Whoever invents or discovers any new and useful process, machine, manufacture, or composition of matter, or any new and useful improvement thereof, may obtain a patent therefor, subject to the conditions and requirements of this title.

4. Claims 1-12 are rejected under 35 U.S.C. 101 because the claimed invention is directed to non-statutory subject matter. The claims are drawn to methods of selecting probe sets. The claimed selection does not produce a physical transformation or produce a tangible result. The claims are methods for selecting probes, but the selecting does not transform the probes or

Art Unit: 1634

physically alter the probes so as to produce a tangible result or change to the probes. The methods merely select or name probes for future use, but the future use is not encompassed within the method steps of the claims.

The courts have stated that “While a scientific truth, or the mathematical expression of it, is not patentable invention, a novel and useful structure created with the aid of knowledge of scientific truth may be.”; Warmerdam, 33 F.3d at 1360, 31 USPQ2d at 1759 (“steps of locating’ a medial axis, and creating’ a bubble hierarchy . . . describe nothing more than the manipulation of basic mathematical constructs, the paradigmatic abstract idea”) (see MPEM § 2106 IV).

The courts have further stated that manipulation of abstract concepts or ideas constitute non-statutory subject matter.

If the “acts” of a claimed process manipulate only numbers, abstract concepts or ideas, or signals representing any of the foregoing, the acts are not being applied to appropriate subject matter. Schrader, 22 F.3d at 294-95, 30 USPQ2d at 1458-59. Thus, a process consisting solely of mathematical operations, i.e., converting one set of numbers into another set of numbers, does not manipulate appropriate subject matter and thus cannot constitute a statutory process.

In practical terms, claims define nonstatutory processes if they:

- consist solely of mathematical operations without some claimed practical application

(i.e., executing a “mathematical algorithm”); or

- **simply manipulate abstract ideas**, e.g. a bid (Schrader, 22 F.3d at 293-94, 30 USPQ2d at 1458-59) or a bubble hierarchy (Warmerdam, 33 F.3d at 1360, 31 USPQ2d at 1759), without some claimed practical application.

(see MPEM § 2106 IV (B) (1)).

It is further noted that In re Schrader states: “the grouping or regrouping of bids cannot constitute a physical change, effect or result”.... “The only physical effect or result which is required by the claim is the entering of bids in a “record,” a step that can be accomplished simply by writing the bids on a piece of paper or chalkboard. For purposes of Section 101,

Art Unit: 1634

such activity is indistinguishable from the data gathering steps which we said in *In re Grams*, 888 F.2d 835, 12 USPQ 2d 1924 (Fed. Cir. 1989), were insufficient to impart patentability to a claimed involving the solving of a mathematical algorithm". Therefore, the courts have stated that manipulation of concepts or signals representing the concepts, without physical manipulation, is indistinguishable from data gathering and insufficient to impart patentability. Hence, the instant claims drawn to selecting without any physical manipulation is non-statutory subject matter.

Claim Rejections - 35 USC § 102

5. The following is a quotation of the appropriate paragraphs of 35 U.S.C. 102 that form the basis for the rejections under this section made in this Office action:

A person shall be entitled to a patent unless –

(a) the invention was known or used by others in this country, or patented or described in a printed publication in this or a foreign country, before the invention thereof by the applicant for a patent.

(b) the invention was patented or described in a printed publication in this or a foreign country or in public use or on sale in this country, more than one year prior to the date of application for patent in the United States.

(e) the invention was described in (1) an application for patent, published under section 122(b), by another filed in the United States before the invention by the applicant for patent or (2) a patent granted on an application for patent by another filed in the United States before the invention by the applicant for patent, except that an international application filed under the treaty defined in section 351(a) shall have the effects for purposes of this subsection of an application filed in the United States only if the international application designated the United States and was published under Article 21(2) of such treaty in the English language.

6. Claims 1-24 are rejected under 35 U.S.C. 102(a) and (e) as being anticipated by Chaganti et al (U.S. Patent No. 6,309,860, issued 30 October 2001).

Regarding Claim 1, Chaganti et al disclose a method of designing nucleic acid probes to a transcription cluster (i.e. BLC-8 transcripts) comprising selecting a first set of probes comprising at least one probe (i.e. probes A & C) targeting a first region immediately upstream

Art Unit: 1634

of a first polyadenylation site in a transcription center and selecting a second probe comprising at least one probe (i.e. multiple copies of probe B) targeting a second region immediately upstream of a second polyadenylation site wherein the first and second regions are different (Column 12, lines 32-51 and Fig. 6).

Regarding Claim 2, Chaganti et al disclose the method wherein the polyadenylation sites are alternative polyadenylation sites (Column 12, lines 32-35).

Regarding Claim 3, Chaganti et al disclose the method wherein the first polyadenylation site is a putative polyadenylation site (Column 12, lines 32-35).

Regarding Claim 4, Chaganti et al disclose the method wherein the first region is within 800 bases upstream of the first polyadenylation site and the second region is within 800 bases upstream of the second polyadenylation site (Column 12, lines 32-51 and Fig. 6).

Regarding Claim 5, Chaganti et al disclose the method wherein the first region is within 600 bases upstream of the first polyadenylation site and the second region is within 600 bases upstream of the second polyadenylation site (Column 12, lines 32-51 and Fig. 6).

Regarding Claim 6, Chaganti et al disclose the method wherein the target nucleic acid represents a cluster of transcript sequences including RNA and ESTs i.e. both types of bcl-8 transcripts (Column 12, lines 32-35).

Regarding Claim 7, Chaganti et al disclose the method wherein the first polyadenylation site is a full length mRNA and the first set of probes (i.e. A & C) are selected to target the full length mRNA as an exemplar sequence of the cluster (Column 12, lines 32-51 and Fig. 6).

Regarding Claim 8, Chaganti et al disclose the method wherein the first polyadenylation site is shared by a stack of sequences in the cluster (i.e. sequences expressed in different tissues) and the probes are selected to target the consensus sequence of the cluster and wherein the stack of sequences comprises at least two sequences i.e. when the internal polyadenylation site is the named the "first polyadenylation site", the first probe set comprises

Art Unit: 1634

A & B. Probe set A & B target the consensus sequences comprising at least two sequences (Fig. 6).

Regarding Claims 9-10, Chaganti et al disclose the method wherein the stack of sequences comprises at least 8 sequences i.e. at least 8 different tissues (Fig. 6).

Regarding Claims 11-12, Chaganti et al disclose the method wherein the probe sets comprise at least 10 probes i.e. multiple copies of probes A & B or A & C (Column 12, lines 32-51 and Fig. 6).

Regarding Claim 13, Chaganti et al disclose an array of nucleic acid probes comprising at least one probe (i.e. probes A & C) targeting a first region immediately upstream of a first polyadenylation site in a transcription center (i.e. BCL-8) and at least one second probe (i.e. multiple copies of probe B) targeting a second region immediately upstream of a second polyadenylation site wherein the first and second regions are different (Column 12, lines 32-51 and Fig. 6). It is noted that the claims are drawn to a nucleic acid probe array. However, the claims are not limited to immobilized or addressable probes. The claims are given the broadest reasonable interpretation consistent with the broad claim language. As such, the broadly claimed probes encompass the probes of Chaganti et al.

Regarding Claim 14, Chaganti et al disclose the probes wherein the polyadenylation sites are alternative polyadenylation sites (Column 12, lines 32-35).

Regarding Claim 15, Chaganti et al disclose the probes wherein the first polyadenylation site is a putative polyadenylation site (Column 12, lines 32-35).

Regarding Claim 16, Chaganti et al disclose the probes wherein the first region is within 800 bases upstream of the first polyadenylation site and the second region is within 800 bases upstream of the second polyadenylation site (Column 12, lines 32-51 and Fig. 6).

Regarding Claim 17, Chaganti et al disclose the probes wherein the first region is within 600 bases upstream of the first polyadenylation site and the second region is within 600 bases upstream of the second polyadenylation site (Column 12, lines 32-51 and Fig. 6).

Art Unit: 1634

Regarding Claim 18, Chaganti et al disclose the probes wherein the target nucleic acid represents a cluster of transcript sequences including RNA and ESTs i.e. both types of bcl-8 transcripts (Column 12, lines 32-35).

Regarding Claim 19, Chaganti et al disclose the probes wherein the first polyadenylation site is a full length mRNA and the first set of probes (i.e. A & C) are selected to target the full length mRNA as an exemplar sequence of the cluster (Column 12, lines 32-51 and Fig. 6).

Regarding Claim 20, Chaganti et al disclose the probes wherein the first polyadenylation site is shared by a stack of sequences in the cluster (i.e. sequences expressed in different tissues) and the probes are selected to target the consensus sequence of the cluster and wherein the stack of sequences comprises at least two sequences i.e. when the internal polyadenylation site is named the "first polyadenylation site", the first probe set comprises A & B. Probe set A & B target the consensus sequences comprising at least two sequences (Fig. 6).

Regarding Claims 21-22, Chaganti et al disclose the probes wherein the stack of sequences comprises at least 8 sequences i.e. at least 8 different tissues (Fig. 6).

Regarding Claims 22-23, Chaganti et al disclose the probes wherein the probe sets comprise at least 10 probes i.e. multiple copies of probes A & B or A & C (Column 12, lines 32-51 and Fig. 6).

7. Claims 1-24 are rejected under 35 U.S.C. 102(b) as being anticipated by Olsen et al (U.S. Patent No. 5,643,783, issued 1 July 1997).

Regarding Claim 1, Olsen et al disclose a method of designing nucleic acid probes to a transcription cluster (i.e. collagen transcripts) comprising selecting a first set of probes

Art Unit: 1634

comprising at least one probe (i.e. probe A) targeting a first region immediately upstream of a first polyadenylation site in a transcription center and selecting a second probe comprising at least one probe (i.e. probe B) targeting a second region immediately upstream of a second polyadenylation site wherein the first and second regions are different (Column 9, lines 45-59 and Fig. 5).

Regarding Claim 2, Olsen et al disclose the method wherein the polyadenylation sites are alternative polyadenylation sites (Column 4, line 60-Column 5, line 2).

Regarding Claim 3, Olsen et al disclose the method wherein the first polyadenylation site is a putative polyadenylation site (Column 4, line 60-Column 5, line 2).

Regarding Claim 4, Olsen et al disclose the method wherein the first region is within 800 bases upstream of the first polyadenylation site and the second region is within 800 bases upstream of the second polyadenylation site (Fig. 5 and Column 11, lines 10-32).

Regarding Claim 5, Olsen et al disclose the method wherein the first region is within 600 bases upstream of the first polyadenylation site and the second region is within 600 bases upstream of the second polyadenylation site (Column 11, lines 10-32 and Fig. 5).

Regarding Claim 6, Olsen et al disclose the method wherein the target nucleic acid represents a cluster of transcript sequences including RNA and ESTs i.e. collagen transcripts (Column 11, lines 10-32).

Regarding Claim 7, Olsen et al disclose the method wherein the first polyadenylation site is a full length mRNA and the first set of probes (i.e. A) are selected to target the full length mRNA as an exemplar sequence of the cluster (Column 9, lines 45-59; Column 11, lines 10-32 and Fig. 5).

Regarding Claim 8, Olsen et al disclose the method wherein the first polyadenylation site is shared by a stack of sequences in the cluster (i.e. sequences expressed in different tissues) and the probes are selected to target the consensus sequence of the cluster and

Art Unit: 1634

wherein the stack of sequences comprises at least two sequences i.e. different transcripts (Column 9, lines 45-59; Column 11, lines 10-32 and Fig. 5).

Regarding Claims 9-10, Olsen et al disclose the method wherein the stack of sequences comprises at least 8 sequences i.e. at least 8 different tissues (Column 9, lines 45-59; Column 11, lines 10-32 and Fig. 5).

Regarding Claims 11-12, Olsen et al disclose the method wherein the probe sets comprise at least 10 probes i.e. multiple copies of probes A & B (Column 9, lines 45-59; Column 11, lines 10-32 and Fig. 5).

Regarding Claim 13, Olsen et al disclose an array of nucleic acid probes comprising at least one probe (i.e. probe A) targeting a first region immediately upstream of a first polyadenylation site in a transcription center (i.e. collagen transcripts) and selecting a second probe comprising at least one probe (i.e. probe B) targeting a second region immediately upstream of a second polyadenylation site wherein the first and second regions are different (Column 9, lines 45-59 and Fig. 5). It is noted that the claims are drawn to a nucleic acid probe array. However, the claims are not limited to immobilized or addressable probes. The claims are given the broadest reasonable interpretation consistent with the broad claim language. As such, the broadly claimed probes encompass the probes of Olsen et al.

Regarding Claim 14, Olsen et al disclose the method wherein the polyadenylation sites are alternative polyadenylation sites (Column 4, line 60-Column 5, line 2).

Regarding Claim 15, Olsen et al disclose the method wherein the first polyadenylation site is a putative polyadenylation site (Column 4, line 60-Column 5, line 2).

Regarding Claim 16, Olsen et al disclose the method wherein the first region is within 800 bases upstream of the first polyadenylation site and the second region is within 800 bases upstream of the second polyadenylation site (Fig. 5 and Column 11, lines 10-32).

Art Unit: 1634

Regarding Claim 17, Olsen et al disclose the method wherein the first region is within 600 bases upstream of the first polyadenylation site and the second region is within 600 bases upstream of the second polyadenylation site (Column 11, lines 10-32 and Fig. 5).

Regarding Claim 18, Olsen et al disclose the method wherein the target nucleic acid represents a cluster of transcript sequences including RNA and ESTs i.e. collagen transcripts (Column 11, lines 10-32).

Regarding Claim 19, Olsen et al disclose the method wherein the first polyadenylation site is a full length mRNA and the first set of probes (i.e. A) are selected to target the full length mRNA as an exemplar sequence of the cluster (Column 9, lines 45-59; Column 11, lines 10-32 and Fig. 5).

Regarding Claim 20, Olsen et al disclose the method wherein the first polyadenylation site is shared by a stack of sequences in the cluster (i.e. sequences expressed in different tissues) and the probes are selected to target the consensus sequence of the cluster and wherein the stack of sequences comprises at least two sequences i.e. different transcripts (Column 9, lines 45-59; Column 11, lines 10-32 and Fig. 5).

Regarding Claims 21-22, Olsen et al disclose the method wherein the stack of sequences comprises at least 8 sequences i.e. at least 8 different tissues (Column 9, lines 45-59; Column 11, lines 10-32 and Fig. 5).

Regarding Claims 23-24, Olsen et al disclose the method wherein the probe sets comprise at least 10 probes i.e. multiple copies of probes A & B (Column 9, lines 45-59; Column 11, lines 10-32 and Fig. 5).

8. Claims 1-24 are rejected under 35 U.S.C. 102(e) as being anticipated by Beattie et al (U.S. Patent No. 6,268,147, filed 2 November 1999).

Art Unit: 1634

Regarding Claim 1, Beattie et al disclose a method of designing a probe array to target a transcription cluster comprising selection a first probe comprising at least one probe targeting a first region immediately upstream of a first polyadenylation site in a transcription center and selecting a second probe comprising at least one probe targeting a second region immediately upstream of a second polyadenylation site wherein the first and second regions are different i.e. probes of Beattie target regions adjacent to the poly-A tail of mRNA and comprise poly-T sequence plus 1-4 bases comprising a mixture of all 4 bases whereby each different probe will target a different polyadenylation site because the bases adjacent to the poly-T sequence of the probe will recognize different sequences (Column 14, line 59-Column 15, line 15 and Example 14, Column 34, lines 1-58).

Regarding Claim 2, Beattie et al disclose the method wherein the polyadenylation sites are alternative polyadenylation sites (Example 14, Column 34, lines 1-58).

Regarding Claim 3, Beattie et al disclose the method wherein the first polyadenylation site is a putative polyadenylation site (Example 14, Column 34, lines 1-58).

Regarding Claim 4, Beattie et al disclose the method wherein the first region is within 800 bases upstream of the first polyadenylation site and the second region is within 800 bases upstream of the second polyadenylation site i.e. adjacent to the poly-A tail (Example 14; Column 34, lines 1-58).

Regarding Claim 5, Beattie et al disclose the method wherein the first region is within 600 bases upstream of the first polyadenylation site and the second region is within 600 bases upstream of the second polyadenylation site i.e. adjacent to the poly-A tail (Example 14, Column 34, lines 1-58).

Regarding Claim 6, Beattie et al disclose the method wherein the target nucleic acid represents a cluster of transcript sequences including RNA and ESTs (Example 14, Column 34, lines 1-58).

Art Unit: 1634

Regarding Claim 7, Beattie et al disclose the method wherein the first polyadenylation site is a full length mRNA and the first set of probes are selected to target the full length mRNA as an exemplar sequence of the cluster (Column 14, line 59-Column 15, line 15; Example 14, Column 34, lines 1-58; and Column 36, lines 25-30).

Regarding Claim 8, Beattie et al disclose the method wherein the first polyadenylation site is shared by a stack of sequences in the cluster and the probes are selected to target the consensus sequence of the cluster and wherein the stack of sequences comprises at least two sequences cluster (Column 14, line 59-Column 15, line 15; Example 14, Column 34, lines 1-58).

Regarding Claims 9-10, Beattie et al disclose the method wherein the stack of sequences comprises at least 8 sequences i.e. mixture of all 4 bases (Example 14, Column 34, lines 1-58).

Regarding Claims 11-12, Beattie et al disclose the method wherein the probe sets comprise at least 10 probes i.e. mixture of all 4 bases (Example 14, Column 34, lines 1-58 and Fig. 1).

Regarding Claim 13, Beattie et al disclose an array of nucleic acid probes comprising at least one probe targeting a first region immediately upstream of a first polyadenylation site in a transcription center and selecting a second probe comprising at least one probe targeting a second region immediately upstream of a second polyadenylation site wherein the first and second regions are different i.e. probes of Beattie target regions adjacent to the poly-A tail of mRNA and comprise poly-T sequence plus 1-4 bases comprising a mixture of all 4 bases whereby each different probe will target a different polyadenylation site because the bases adjacent to the poly-T sequence of the probe will recognize different sequences (Example 14, Column 34, lines 1-58).

Regarding Claim 14, Beattie et al disclose the array wherein the polyadenylation sites are alternative polyadenylation sites (Example 14, Column 34, lines 1-58).

Art Unit: 1634

Regarding Claim 15, Beattie et al disclose the array wherein the first polyadenylation site is a putative polyadenylation site (Example 14, Column 34, lines 1-58).

Regarding Claim 16, Beattie et al disclose the array wherein the first region is within 800 bases upstream of the first polyadenylation site and the second region is within 800 bases upstream of the second polyadenylation site i.e. adjacent to the poly-A tail (Example 14, Column 34, lines 1-58).

Regarding Claim 17, Beattie et al disclose the array wherein the first region is within 600 bases upstream of the first polyadenylation site and the second region is within 600 bases upstream of the second polyadenylation site i.e. adjacent to the poly-A tail (Example 14, Column 34, lines 1-58).

Regarding Claim 18, Beattie et al disclose the array wherein the target nucleic acid represents a cluster of transcript sequences including RNA and ESTs (Example 14, Column 34, lines 1-58).

Regarding Claim 19, Beattie et al disclose the array wherein the first polyadenylation site is a full length mRNA and the first set of probes are selected to target the full length mRNA as an exemplar sequence of the cluster (Column 14, line 59-Column 15, line 15; Example 14, Column 34, lines 1-58; and Column 36, lines 25-30).

Regarding Claim 20, Beattie et al disclose the array wherein the first polyadenylation site is shared by a stack of sequences in the cluster and the probes are selected to target the consensus sequence of the cluster and wherein the stack of sequences comprises at least two sequences cluster (Column 14, line 59-Column 15, line 15; Example 14, Column 34, lines 1-58).

Regarding Claims 21-22, Beattie et al disclose the array wherein the stack of sequences comprises at least 8 sequences i.e. mixture of all 4 bases (Example 14, Column 34, lines 1-58).

Art Unit: 1634

Regarding Claims 23-24, Beattie et al disclose the array wherein the probe sets comprise at least 10 probes i.e. mixture of all 4 bases (Example 14, Column 34, lines 1-58 and Fig. 1).

Claim Rejections - 35 USC § 103

9. The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

10. Claims 25 and 26 are rejected under 35 U.S.C. 103(a) as being unpatentable over Beattie et al (U.S. Patent No. 6,268,147, filed 2 November 1999) in view of Lockhart et al (U.S. Patent No. 5,556,752, filed 24 October 1994).

Regarding Claims 25-26, Beattie et al teaches the array of nucleic acid probes comprising at least one probe targeting a first region immediately upstream of a first polyadenylation site in a transcription center and selecting a second probe comprising at least one probe targeting a second region immediately upstream of a second polyadenylation site wherein the first and second regions are different (Example 14, Column 34, lines 1-58) but they are silent regarding the probe density. However, probe density of greater than 400 different probes/cm² (Claim 25) and greater than 1000 different probes/cm² (Claim 26) were well known in the art at the time the claimed invention was made as taught by Lockhart et al who specifically teach polyadenylation site-specific probes immobilized at a density greater than 1000 different probes/cm² (Column 10, lines 45-55 and Column 13, lines 19-24). It would have been obvious to one of ordinary skill in the art at the time the claimed invention was made to apply

Art Unit: 1634

the probe density of Lockhart et al to the probe array of Beattie to thereby analyze the genomes of complete organisms on a single support as desired by Beattie (Column 34, lines 50-58).

Double Patenting

11. The nonstatutory double patenting rejection is based on a judicially created doctrine grounded in public policy (a policy reflected in the statute) so as to prevent the unjustified or improper timewise extension of the “right to exclude” granted by a patent and to prevent possible harassment by multiple assignees. A nonstatutory obviousness-type double patenting rejection is appropriate where the conflicting claims are not identical, but at least one examined application claim is not patentably distinct from the reference claim(s) because the examined application claim is either anticipated by, or would have been obvious over, the reference claim(s). See, e.g., *In re Berg*, 140 F.3d 1428, 46 USPQ2d 1226 (Fed. Cir. 1998); *In re Goodman*, 11 F.3d 1046, 29 USPQ2d 2010 (Fed. Cir. 1993); *In re Longi*, 759 F.2d 887, 225 USPQ 645 (Fed. Cir. 1985); *In re Van Ornum*, 686 F.2d 937, 214 USPQ 761 (CCPA 1982); *In re Vogel*, 422 F.2d 438, 164 USPQ 619 (CCPA 1970); and *In re Thorington*, 418 F.2d 528, 163 USPQ 644 (CCPA 1969).

A timely filed terminal disclaimer in compliance with 37 CFR 1.321(c) or 1.321(d) may be used to overcome an actual or provisional rejection based on a nonstatutory double patenting ground provided the conflicting application or patent either is shown to be commonly owned with this application, or claims an invention made as a result of activities undertaken within the scope of a joint research agreement.

Effective January 1, 1994, a registered attorney or agent of record may sign a terminal disclaimer. A terminal disclaimer signed by the assignee must fully comply with 37 CFR 3.73(b).

12. Claims 1-12 are rejected on the ground of nonstatutory obviousness-type double patenting as being unpatentable over claims 1-2, 4-11, 13-20 and 22-30 of allowed Application No. 09/745,965. Although the conflicting claims are not identical, they are not patentably distinct from each other because both sets of claims are drawn to methods for selecting probes. The claim sets differ in that the allowed methods utilize a computer and algorithms for probe selection. However, the instant claim language “comprising” encompasses the use of a computer and additional calculations recited in the patented methods.

Art Unit: 1634

13. Claims 1-26 are provisionally rejected on the ground of nonstatutory obviousness-type double patenting as being unpatentable over claims 13-17 of copending Application No.

11/121,849. Although the conflicting claims are not identical, they are not patentably distinct from each other because both sets of claims are drawn to probe arrays and methods for their design. The claim sets merely differ in that the instantly claimed probes are designed to have a target region within 600 bases upstream of the poly A site while the '849 is drawn to target regions within 300 bases of the poly A site.

The courts have stated where the claimed ranges “overlap or lie inside the ranges disclose by the prior art” and even when the claimed ranges and prior art ranges do not overlap but are closed enough that one skilled in the art would have expected them to have similar properties, a *prima facie* case of obviousness exists (see *In re Wertheim*, 541 F.2d 257, 191 USPQ 90 (CCPA 1976); *In re Woodruff*, 919 F.2d 1575, 16 USPQ2d 1934 (Fed. Cir. 1990); *Titanium Metals Corp. of America v. Banner*, 778 F.2d 775, 227 USPQ 773 (Fed. Cir. 1985) (see MPEP, 2144.05 I.).

The instantly claimed region overlaps with the region recited in the '849 claims such that one of ordinary skill would have expected the probes to have similar functionality. Therefore, the instantly claimed ranges are obvious variation of the '849 range.

This is a provisional obviousness-type double patenting rejection because the conflicting claims have not in fact been patented.

Conclusion


14. No claim is allowed.

Art Unit: 1634

15. Any inquiry concerning this communication or earlier communications from the examiner should be directed to BJ Forman whose telephone number is (571) 272-0741. The examiner can normally be reached on 6:00 TO 3:30.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Ram Shukla can be reached on (571) 272-0735. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

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Art Unit 1634

3 July 2006